PHOSPHATE ACCUMULATION USING MgO-Fe₃O₄/AGAROSE/OXALIC ACID GEL AS A DIFFUSIVE GRADIENT IN THIN FILMS (DGT) BINDING LAYER

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Abstract: Monitoring the phosphate concentration in water is critical because excessive phosphate can lead to the death of aquatic organisms. Phosphate can be monitored via a passive sampler called the diffusion gradient in thin films (DGT). This study combines MgO and Fe₃O₄ impregnated in agarose crosslinked with oxalic acid to accumulate phosphate from the solution as a DGT binding gel. The parameters observed in this study were the MgO/Fe₃O₄ mass ratio (1:3, 1:1, 3:1), accumulation time (20, 40, 60, 120, 240, and 1440 min), phosphate concentration (0.2, 0.4, 0.6, 0.8, and 1 mg/L) and pH (4, 5, 6, 7, and 8) for phosphate accumulation. The XRD patterns confirmed that the adsorbents used were MgO and Fe₃O₄. SEM analysis revealed that the gel had an average pore size of 31.78 µm, and the adsorbents were evenly distributed. Gels with a MgO/Fe₃O₄ mass ratio of 3:1 can adsorb up to 97.19 \pm 0.36% of the phosphate. The phosphate accumulation reached an optimum after a minimum adsorption time of 4 h and when the phosphate concentration in the solution was 0.4 mg/L. The pH of the solution had no significant effect on phosphate accumulation. This study revealed that MgO-Fe₃O₄/agarose/oxalic acid gel is an excellent binding gel for the accumulation of phosphate from water.

Keywords: DGT; MgO; Fe₃O₄; agarose; oxalic acid

Abstrak: Pemantauan fosfat dalam perairan sangat penting karena kadar fosfat berlebih dapat mengakibatkan eutrofikasi yang berujung pada kematian organisme air. Pemantauan kadar fosfat dalam perairan dapat menggunakan *passive sampler* yang dinamakan *Diffusive Gradient in Thin Films* (DGT). Penelitian ini mengombinasikan MgO dan Fe₃O₄ terimpregnasi dalam gel agarosa terikat silang asam oksalat untuk mengakumulasi fosfat dari larutan sebagai gel pengikat DGT. Parameter-parameter yang diamati pada penelitian ini adalah pengaruh rasio massa MgO/Fe₃O₄ (1:3; 1:1; 3:1), waktu akumulasi (20; 40; 60; 120; 240; dan 1440 menit), konsentrasi fosfat dalam larutan (0,2; 0,4; 0,6; 0,8; dan 1 mg/L) dan pH larutan (4; 5; 6; 7; dan 8) terhadap akumulasi

fosfat. Hasil penelitian menunjukkan bahwa puncak-puncak yang teramati pada XRD mengonfirmasi bahwa adsorben yang digunakan adalah MgO dan Fe₃O₄. Hasil SEM menunjukkan bahwa gel memiliki ukuran pori rata-rata sebesar 31,78 μ m dan adsorben terdistribusi merata. Gel dengan rasio massa MgO/Fe₃O₄ sebesar 3:1 dapat mengadsorpsi fosfat hingga 97,19 \pm 0,36%. Akumulasi fosfat mencapai optimum setelah adsorpsi minimal selama 4 jam dan ketika konsentrasi fosfat dalam larutan sebesar 0,4 mg/L. pH larutan tidak berpengaruh signifikan terhadap akumulasi fosfat. Penelitian ini menunjukkan bahwa gel MgO-Fe₃O₄/agarosa/asam oksalat dapat mengakumulasi fosfat dari perairan dengan sangat baik.

Kata kunci: fosfat; MgO; Fe₃O₄; agarosa; asam oksalat

INTRODUCTION

Phosphate (PO4³⁻) is an essential chemical for natural biological life. Agriculture uses phosphate as a fertilizer (Puspitasari et al., 2018; Sumbayak & Gultom, 2020). However, excess phosphate can lead to eutrophication, which results in excessive zooplankton growth and proliferation (Badamasi et al., 2019).

According to the Government Regulation of the Republic of Indonesia Number 22 of 2021, phosphate is one of the factors contributing to water pollution in irrigation water and must not exceed 0.1 mg/L (Pemerintah Republik Indonesia, 2022). Thus, monitoring phosphate levels in water is critical for preventing water contamination.

The grab sampling method can determine phosphorus levels in water (Udianto et al., 2022). However, sample analysis in the laboratory using the grab sampling method still requires analyte preconcentration, which might lead to errors in measurement. The diffusive gradient in thin films (DGTs) is a great and straightforward in situ measurement technique (Zheng et al., 2023). It is frequently used to assess aquatic species such as phosphorus (P), sulfur (S), arsenic (As), and metals. A DGT passive sampler consists of a binding layer, a diffusive layer, and a filter membrane (Li et al., 2019; Yabuki et al., 2014). Various phosphate binder layers have been utilized, including zirconium oxide, magnetite, ferihydrite, and Mg(OH)₂ (Maimulyanti et al., 2018; Sun et al., 2013; F. Xie et al., 2020; Y. Zhang et al., 2018).

The use of magnetite as a binding agent is based on its high affinity for phosphate ions (Shahid et al., 2019). Similar research on the use of Fe₃O₄ as a binding agent in DGTs was conducted by Zhang (Y. Zhang et al., 2018). Fe₃O₄ was used as a suspension. However, owing to the high affinity of Fe₃O₄ for phosphate ions, the phosphate concentration measured via DGT was three times greater than the actual phosphate concentration. Therefore, the use of Fe₃O₄ to adsorb phosphate in DGTs must be reduced and replaced with another binding agent. Mg-based adsorbents are one of the available options. According to (F. Xie et al., 2020), Mg(OH)₂ adsorbs up to 21.37 μ g of phosphate per disc binding gel DGT.

The existing studies only used a single binding agent to adsorb the analyte. There has never been a study of DGT-binding gels that use two binding agents to adsorb Combining phosphate. а Mg-based adsorbent with Fe₃O₄ is expected to reduce the high affinity of Fe₃O₄ in adsorbing phosphate without reducing the excellent phosphate adsorption percentage (~100%). The DGT-binding gel used a hydrogel as the binding agent matrix. Polyacrylamide and agarose are commonly used as hydrogels for DGTs (H. Zhang & Davison, 1999). However, unlike agarose, polyacrylamide, is an environmentally friendly hydrogel (Date et al., 2020). This biopolymer can form crosslinks with other compounds because its structure is rich in hydroxyl groups (Jiang et al., 2023; Zucca et al., 2016). It is also safer to prepare and can withstand a pH range of 0-14 (Abolghasemi et al., 2016). The pore size can be adjusted by adjusting the crosslinker concentration (Popescu et al., 2022). Adding crosslinkers also increases the mechanical strength of hydrogels (Zheng et al., 2023).

Oxalic acid is an environmentally friendly dicarboxylate group that can be used as a crosslinker. The structure of oxalic acid consists of two –COOH groups that can form ionic interactions with the helix bonds of biopolymer hydrogel groups. Oxalic acid can form interconnected pores and reduce the pore size of a hydrogel (Popescu et al., 2022).

The ability of the DGT-binding gel to adsorb phosphate must be considered. Several factors can affect phosphate adsorption, such as the mass ratio of the binding agent, adsorption time, pH, and initial concentration of the solution. Therefore, this study developed and optimized the adsorption of phosphate using MgO-Fe₃O₄/agarose/oxalic acid gel.

METHOD

Materials and Apparatus

The chemicals used include FeSO₄.7H₂O (p.a.), FeCl₃.6H₂O (p.a.), 25% NH₄OH solution (p.a.), agarose (p.a.), oxalic acid (p.a.), MgO (p.a.), KH₂PO₄ (p.a.), ammonium heptamolybdate (p.a.), SnCl₂ (p.a.), H₂SO₄ 98% (p.a.), glycerol (p.a.), phenolphthalein (PP) indicator, and HNO₃ solution.

The apparatus used to support the research included glassware, universal pH, micropipettes, circular molds with a diameter of 2.5 cm, gel molds (Figure 1), a UV–Vis spectrophotometer (Thermo Scientific Genesys 10S), X-ray diffraction (PANalytical X'Pert PRO), and scanning electron microscopy (FEI Inspect S50).



Figure 1. The gel mold consists of two pieces of glass (16 x 10 cm). A Teflon spacer formed like the letter U was placed in the middle of the mold. The two pieces of glass and the spacer were clamp with a clip.

Synthesis of Fe₃O₄

FeSO₄.7H₂O (2.78 grams) and FeCl₃.6H₂O (5.41 grams) were put into a glass beaker and dissolved in 50 mL of distilled water with a magnetic stirrer for approximately 10 minutes until homogeneous. Moreover, 3 M NH4OH was added to the solution using a syringe pump at 50 mL/hour until the pH reached 10. The precipitate was washed with distilled water until it reached pH 7 and dried in an oven at 60 °C for 1.5 hours.

Synthesis of MgO-Fe₃O₄/Agarose/Oxalic Acid Gel

Oxalic acid (0.015 grams) was dissolved in 3 mL of distilled water. The solution was

added to a glass beaker containing MgO and transferred to another glass beaker containing Fe₃O₄. The mixture was stirred until homogenous.

Distilled water (7 mL) was added to 0.150 grams of agarose and stirred until the agarose completely dissolved. If the agarose did not dissolve completely, the mixture was heated on a hot plate until a clear, colorless solution formed. The agarose mixture was poured into a beaker containing an oxalic acid-MgO-Fe₃O₄ mixture when the temperature of the agarose mixture was 60 °C. The solution was stirred until homogeneous and then pipetted into a mold (Figure 1). The mold was placed in an oven at 40 °C for 15 minutes.

The gel was cooled at room temperature for approximately 15 minutes. The gel was cut using a circular mold with a diameter of 2.5 cm, immersed in distilled water, and stored for a minimum of 24 hours before use. The thickness of the gel after storage for 24 hours was measured. Gels with a MgO:Fe₃O₄ mass ratio of 3:1 were characterized by scanning electron microscopy (FEI Inspect S50).

Effect of the MgO/Fe₃O₄ mass ratio on phosphate accumulation

Three gels with different MgO/Fe₃O₄ mass ratios (1:0; 3:1; 1:1; 1:3; and 0:1) were

each put into an Erlenmeyer flask containing 15 mL of 2 mg/L phosphate solution. Phosphate adsorption was performed by shaking the phosphate solution containing the gel for 24 hours on a shaker at 100 rpm. After 24 h, the solution was filtered, and the gel and filtrate were obtained.

Effect of adsorption time on phosphate accumulation

Three gels with the optimum MgO/Fe₃O₄ mass ratio were placed into an Erlenmeyer flask containing 15 mL of 2 mg/L phosphate solution. The gels and the phosphate solution were shaken for 20, 40, 60, 120, 240, and 1440 minutes at 100 rpm. The solution was filtered to obtain the gel and filtrate.

Effect of Phosphate Concentration on Phosphate Accumulation

Three gels with the optimum MgO/Fe₃O₄ mass ratio were put into an Erlenmeyer flask containing 15 mL of phosphate solution at concentrations of 0.2, 0.4, 0.6, 0.8, and 1 mg/L and then shaken for 24 hours at 100 rpm. Filtration was used to separate the gel and filtrate.

Effect of pH on Phosphate Accumulation

Three gels with the optimum MgO/Fe₃O₄ mass ratio were put into an Erlenmeyer

flask containing 15 mL of 0.4 mg/L phosphate solution with a pH of 4-8 (adjusted with HNO₃ and NaOH) and then shaken at 100 rpm for 24 hours. The solution was filtered to obtain a gel and filtrate.

Data analysis

The crystallite diameter of the binding agent was calculated using the Debye– Scherrer equation, as shown below:

$$D = \frac{\kappa\lambda}{\beta\cos\theta}$$
(2)

 β is the FWHM (full width at half maximum or half width) (in radians), θ is the position of the maximum diffraction peak, K is Scherrer's constant of 0.9, and λ is the X-ray wavelength (1.5406 Å or 0.15406 nm).

The initial phosphate concentration before and after adsorption was analyzed using the Stannous Chloride method. The phosphate concentration was converted into the phosphate mass before (m_o) and after adsorption (m_t) . The phosphate accumulation (%) was calculated using the equation below:

Phosphate accumulation (%) = $\frac{m_o - m_t}{m_o} \times 100\%$ (1) The phosphate accumulation obtained for each variation was analyzed using oneway ANOVA followed by Tukey's test at the 95% confidence level (p ≤ 0.05).

RESULTS AND DISCUSSION

Characterization of the MgO-Fe₃O₄/Agarose/Oxalic Acid Gel

synthesis of FeSO₄.7H₂O The and $Fe^{2+}:Fe^{3+} =$ FeCl³.6H₂O (mole 1:2)produced a black precipitate. The precipitate was washed to pH 7, dried, and characterized using XRD. Figure 2 shows that the precipitate was Fe₃O₄ because there were peaks at 30° , 35° , 43° , 54° , 57° , and 63° , which is the typical 2θ of Fe₃O₄. These peaks indicated the (220), (311), (400), (422), (511), and (440) fields (Bertolucci et al., 2015). The Debye-Scherrer equation calculation revealed that the average diameter of the Fe₃O₄ crystallites (D) was 19.08 nm.

Figure 3 shows the XRD characterization of the MgO used in this study. The XRD pattern indicated typical MgO angles at 36°, 42°, 62°, 74°, and 78°, which correspond to the (111), (200), (220), (311), and (222) planes, respectively, according to the reference standard document (JCPDS 98-015-9369) (Demirci et al., 2021; Salman et al., 2021). The average crystallite diameter of MgO (D) determined using the Debye–Scherrer equation was 71.76 nm.



Figure 2. XRD pattern of Fe₃O₄



Figure 3. XRD pattern of MgO

The MgO and Fe₃O₄ were impregnated into agarose crosslinked with oxalic acid gel and cut into a round shape with a diameter of 2.5 cm. The color of the gel was gray due to a mixture of white MgO and black Fe₃O₄. It had a thickness of 0.5 mm. A binding gel of this size can be inserted into the DGT passive sampler with a total layer thickness of 1.34 mm (Davison, 2016).

Dinira et al. (2024) crosslinked agarose with oxalic acid. The IR spectrum of agarose-oxalic acid revealed a new peak at 1727 cm⁻¹ that was not present in the IR spectrum of agarose. This suggests that agarose was successfully crosslinked with oxalic acid via an esterification reaction.

The gel was stored in distilled water for 24 hours. During storage, the distilled water was changed three times. The MgO/Fe₃O₄/agarose oxalic acid gel was characterized using SEM to evaluate the pore size and binding agent distribution of the gel. Figure 4 shows the surface of the gel. The white areas indicate that MgO and Fe₃O₄ were found and spread well in the gel matrix. The average pore size of the gel was 31.78 µm.



Figure 4. SEM image of MgO/Fe₃O₄ 3:1 impregnated in agarose-oxalic acid gel

Effect of the MgO/Fe₃O₄ mass ratio on the phosphate accumulation in the MgO/Fe₃O₄/agarose/oxalic acid gel

The accumulation of phosphate in MgO/Fe₃O₄/agarose/oxalic acid gels of various mass ratios is shown in Figure 5. All the gels accumulated more than 90% phosphate. The highest phosphate accumulation was obtained when the gel

containing only Fe₃O₄ was used (98.09 \pm 0.44%).

The pH of the phosphate solution used in this study was 6.13. At pH values greater than 4.6, Fe₃O₄ is negatively charged (Zhu et al., 2015) and repels phosphate, decreasing its accumulation. The high degree of phosphate accumulation is due to the high affinity of Fe^{3+} in Fe_3O_4 for phosphate (Senn et al., 2015). The high phosphate accumulation in the gel contained only MgO (97.5 \pm 0.35%), which was attributed to the positively charged MgO surface at pH < 9.2 (Ismail et al., 2022). The positively charged MgO surface can adsorb phosphate via electrostatic interactions (Wang et al., 2024).



Figure 5. Effect of binding agent mass ratio on phosphate accumulation

According to the statistical test, phosphate accumulation in the gel was not significantly different from that in the gel containing only MgO or the gel containing MgO/Fe₃O₄ at a 3:1 ratio. (97.19 \pm

0.36%). This result demonstrates that when Fe₃O₄ is replaced with MgO, the gel can still adsorb phosphate effectively. However, iron oxide has a strong ability to attract phosphate; thus, it is still required to adsorb phosphate (Z. Zhang et al., 2020). As a result, the MgO/Fe₃O₄/agarose/oxalic acid gel at a 3:1 ratio was used to determine the effect of adsorption time.

Effect of Adsorption Time on Phosphate Accumulation in MgO/Fe₃O₄/Agarose/oxalic acid gels

Figure 6 shows phosphate accumulation on a MgO/Fe₃O₄/agarose/oxalic acid gel with an optimal MgO/Fe₃O₄ mass ratio over time. Phosphate accumulated quickly in the first 60 min (92.21 \pm 6.49%). Figure 5 also shows that as the contact time between the binding gel and the phosphate solution increased, the amount of phosphate adsorbed increased until the optimal conditions were reached.

The study revealed that phosphate adsorption was most effective after 4 h (98.40 \pm 0.05%) and remained constant for 24 h (98.73 \pm 0.95%). This phenomenon suggests that equilibrium has been attained after at least 4 hours of adsorption (Saleh, 2022). The gel saturation in this study was achieved faster than that of the binding gel, which only used Mg(OH)₂. The accumulation of phosphate by $Mg(OH)_2$ slowed after 20 hours of adsorption (F. Xie et al., 2020).

Statistical tests revealed a p value (0.00) < $\alpha = 0.05$, indicating a significant effect of adsorption time phosphate on accumulation in the gel. However, the statistical test revealed no significant difference between 60 and 1440 minutes. There were some fluctuations between 60 and 240 minutes. As a result, 4 hours was determined to be the minimum period for adsorption phosphate by the MgO/Fe₃O₄/agarose/oxalic acid gel.



Figure 6. Effect of adsorption time on phosphate accumulation

Effect of Initial Solution Concentration on Phosphate Accumulation in MgO/Fe₃O₄/Agarose/Acid Oxalate Gels

The effect of the initial solution concentration on the accumulation of phosphate in the gel is shown in Figure 7. The optimum phosphate accumulation was reached when a phosphate solution with a concentration of 0.4 mg/L was used (95.54 \pm 1.99%). The degree of phosphate accumulation tended to be constant until the concentration reached 1 mg/L (93.68 \pm 2.12%). The greater the amount of phosphate available in the solution is, the greater the degree of phosphate adsorption by the gel until the optimum conditions are reached. A higher concentration of phosphate solution in the solution provides a stronger driving force for the migration of phosphate from the solution to the gel surface (Du et al., 2022) so that the active sites of the gel can adsorb more phosphate. However, the amount of phosphate accumulated tends to be constant after the optimum conditions are reached because the active sites of the gel are saturated (Saefumillah & H, 2015).



Figure 7. Effect of initial concentration of phosphate in solution on phosphate accumulation

Statistical tests revealed a p value $(0.00) < \alpha = 0.05$. It can be concluded that the initial concentration of a solution significantly affects phosphate

accumulation. The amount of phosphate accumulation decreased when the gel adsorbed phosphate from a solution with a one mg/L phosphate concentration. However, statistical tests revealed no significant difference in the amount of phosphate accumulated from phosphate solutions with concentrations ranging from 0.4 to 1 mg/L. Therefore, a phosphate concentration of 0.4 mg/L was used as the solution for the next variation.

Effect of Solution pH on Phosphate Accumulation in MgO/Fe₃O₄/Agarose/Oxalic Acid Gel

Figure 8 shows the effect of solution pH on phosphate accumulation in the gel. At pH 4-8, phosphate accumulation is relatively constant (approximately 90%). A pH of 6 resulted in maximum phosphate accumulation (93.35 \pm 1.78%). At pH 6, the phosphate in the solution is in the form of $H_2PO_4^-$ (J. Xie et al., 2014), whereas MgO, an adsorbent with a relatively high mass, is positively charged, attracting $H_2PO_4^-$ ions to the gel. At pH 8, phosphate in solution begins to appear as HPO_4^{2-} ; at higher pH values, the surface of Fe₃O₄ gradually converts to FeO⁻ (Karunanayake et al., 2019). The negative charges of HPO₄²⁻ and FeO⁻ resist each other, reducing phosphate accumulation.

Statistical tests revealed no significant effect of pH on phosphate accumulation (p value = 0.426, α = 0.05). This suggests that the gel can adsorb phosphate in waters with a pH of 4–8. The gel developed in this study has a lower pH range than the DGT binding gel, which contains only Mg(OH)₂. The Mg(OH)₂ binding gel can be deployed at pH 2–10 (F. Xie et al., 2020).



REFERENCES

Abolghasemi, M. M., Sobhi, M., & Piryaei, M. (2016). Preparation of a novel green optical pH sensor based on immobilization of red grape extract on bioorganic agarose membrane. Sensors and Actuators, B: Chemical, 224, 391–395. https://doi.org/10.1016/j.snb.2015.10. 038

CONCLUSION

The critical finding of this study is that the MgO-Fe₃O₄/agarose/oxalic acid gel, as a binding gel, accumulates up to 100% phosphate. The mass ratio of MgO:Fe₃O₄, adsorption time, and initial concentration of the phosphate solution significantly affect the accumulation of phosphate by the MgO-Fe₃O₄/agarose/oxalic acid gel. Optimal phosphate adsorption occurred when a gel with a MgO/Fe₃O₄ mass ratio of 3:1, a minimum adsorption time of 4 hours, and a phosphate content of 0.4 mg/L was used. Since phosphate accumulation in phosphate solutions at pH 4-8 tends to be constant, the gel can adsorb phosphate in water within this pH range. This study revealed that the MgO-Fe₃O₄/agarose/oxalic acid gel is an excellent binding gel for the accumulation of phosphate from water.

- Badamasi, H., Yaro, M. N., Ibrahim, A.,
 & Bashir, I. A. (2019). Impacts of Phosphates on Water Quality and Aquatic Life. *Chemical Research Journal*, 4(3), 124–133.
- Bertolucci, E., Galletti, A. M. R.,Antonetti, C., Marracci, M., Tellini,B., Piccinelli, F., & Visone, C.(2015). Chemical and magnetic

properties characterization of magnetic nanoparticles. *Conference Record - IEEE Instrumentation and Measurement Technology Conference, 2015-July,* 1492–1496. https://doi.org/10.1109/I2MTC.2015. 7151498

- Date, P., Tanwar, A., Ladage, P., Kodam,
 K. M., & Ottoor, D. (2020).
 Biodegradable and biocompatible agarose–poly (vinyl alcohol) hydrogel for the in vitro investigation of ibuprofen release. *Chemical Papers*, 74(6), 1965–1978.
 https://doi.org/10.1007/s11696-019-01046-8
 - Davison, W. (2016). Diffusive Gradients in Thin-Films for Environmental Measurements. Cambridge University Press.
 - Demirci, S., Yildirim, B. K., Tünçay, M.
 M., Kaya, N., & Güllüoğlu, A. N.
 (2021). Synthesis, characterization, thermal, and antibacterial activity studies on MgO powders. *Journal of Sol-Gel Science and Technology*, 99(3), 576–588. https://doi.org/10.1007/s10971-021-05609-8
 - Dinira, L., Rumhayati, B., Andayani, U.,& Mardiana, D. (2024). Effect of casting temperature on the swelling degree and water content of agarose

crosslinked with oxalic acid as DGT diffusion layer. *AIP Conference Proceedings*, *3068*(1). https://doi.org/10.1063/5.0202031

- Du, M., Zhang, Y., Wang, Z., Lv, M., Tang, A., Yu, Y., Qu, X., Chen, Z., Wen, Q., & Li, A. (2022). Insight into the synthesis and adsorption mechanism of adsorbents for efficient phosphate removal: Exploration from synthesis to modification. *Chemical Engineering Journal*, 442, 136147. https://doi.org/10.1016/j.cej.2022.136 147
- Ismail, M., Jobara, A., Bekouche, H., Abd Allateef, M., Ben Aissa, M. A., & Modwi, A. (2022). Impact of Cu Ions removal onto MgO nanostructures: adsorption capacity and mechanism. *Journal of Materials Science: Materials in Electronics*, 33(15), 12500–12512. https://doi.org/10.1007/s10854-022-

08207-8

- Jiang, F., Xu, X. W., Chen, F. Q., Weng,
 H. F., Chen, J., Ru, Y., Xiao, Q., &
 Xiao, A. F. (2023). Extraction,
 Modification and Biomedical
 Application of Agarose Hydrogels: A
 Review. *Marine Drugs*, 21(5), 299.
 https://doi.org/10.3390/md21050299
- Karunanayake, A. G., Navarathna, C. M., Gunatilake, S. R., Crowley, M.,

Anderson, R., Mohan, D., Perez, F., Pittman, C. U., & Mlsna, T. (2019). Fe3O4 Nanoparticles Dispersed on Douglas Fir Biochar for Phosphate Sorption. *ACS Applied Nano Materials*, 2(6), 3467–3479. https://doi.org/10.1021/acsanm.9b004 30

- Li, C., Ding, S., Yang, L., Wang, Y., Ren, M., Chen, M., Fan, X., & Lichtfouse, E. (2019). Diffusive gradients in thin films: devices, materials and applications. *Environmental Chemistry Letters*, 17(2), 801–831. https://doi.org/10.1007/s10311-018-00839-9
- Maimulyanti, A., Budiawan. Saefumillah, A., & Suseno, H. (2018). Effect of pH and anion interferences on determination of orthophosphate speciation by diffusive gradient in thin film (DGT) technique. Rasayan Journal of Chemistry, 11(3), 1222-1228. https://doi.org/10.31788/RJC.2018.11 33057
- Pemerintah Republik Indonesia. (2022). Peraturan Pemerintah Nomor 22 Tahun 2021 tentang Penyelenggaraan Perlindungan dan Pengelolaan Lingkungan Hidup. In *Peraturan*

Pemerintah. Kementerian Sekretariat Negara Republik Indonesia.

- Popescu, I., Constantin, M., Pelin, I. M., Suflet, D. M., Ichim, D. L., Daraba, O. M., & Fundueanu, G. (2022). Eco-Friendly Synthesized PVA/Chitosan/Oxalic Acid Nanocomposite Hydrogels
 Embedding Silver Nanoparticles as Antibacterial Materials. *Gels*, 8(5). https://doi.org/10.3390/gels8050268
- Puspitasari, H. M., Yunus, A., & Harjoko,
 D. (2018). Dosis Pupuk Fosfat
 Terhadap Pertumbuhan Dan Hasil
 Beberapa Jagung Hibrida. *Agrosains*, 20(2), 34–39.
- Saefumillah, A., & H, R. R. (2015). Pengembangan Metode DGT (Diffusive Gradient In Thin Film) dengan Binding Gel Fe-Al-Oksida dan Pengikat Silang N,N'-Methylenebisacrylamide untuk Penyerapan Fosfat dalam Air. Jurnal Kimia VALENSI. 20-25.https://doi.org/10.15408/jkv.v0i0.359 7
- Saleh, T. A. (2022). Kinetic models and thermodynamics of adsorption processes: classification. *Interface Science and Technology*, 34, 65–97. https://doi.org/10.1016/B978-0-12-849876-7.00003-8

- Salman, K. D., Abbas, H. H., & Aljawad, H. A. (2021). Synthesis and characterization of MgO nanoparticle via microwave and sol-gel methods. of Physics: Journal Conference 1973(1), 012104. Series. https://doi.org/10.1088/1742-6596/1973/1/012104
 - Senn, A. C., Kaegi, R., Hug, S. J., Hering, J. G., Mangold, S., & Voegelin, A. (2015). Composition and structure of Fe(III)-precipitates formed by Fe(II) oxidation in water at near-neutral pH: Interdependent effects of phosphate, silicate and Ca. *Geochimica et Cosmochimica Acta*, 162, 220–246. https://doi.org/10.1016/j.gca.2015.04. 032
 - Shahid, M. K., Kim, Y., & Choi, Y. G. (2019). Adsorption of phosphate on magnetite-enriched particles (MEP) separated from the mill scale. *Frontiers of Environmental Science and Engineering*, 13(5), 71–83. https://doi.org/10.1007/s11783-019-1151-2
 - Sumbayak, R. J., & Gultom, R. R. (2020). Pengaruh Pemberian Pupuk Fosfat dan Pupuk Organik. Jurnal Darma Agung, 2, 253–268.
 - Sun, Q., Chen, Y., Xu, D., Wang, Y., & Ding, S. (2013). Investigation of

potential interferences on the measurement of dissolved reactive phosphate using zirconium oxidebased DGT technique. *Journal of Environmental Sciences (China)*, 25(8), 1592–1600. https://doi.org/10.1016/S1001-0742(12)60140-5

- Udianto, F., Kriswandana, F., & Rachmaniyah. (2022). Pemetaan Kualitas Air Sungai di Kawasan Industri Ngingas Sidoarjo Ditinjau dari Parameter BOD dan TSS Tahun 2021. Jurnal Higiene Sanitasi, 1, 31.
- Wang, C. Y., Zhou, H. D., Wang, Q., Xu,
 B. X., & Zhu, G. (2024). Efficiency and mechanism of phosphate adsorption and desorption of a novel Mg-loaded biochar material. *Environmental Science and Pollution Research International*, 31(3), 4425– 4438. https://doi.org/10.1007/s11356-023-31400-z
- Xie, F., Li, L., Sun, X., Hu, T., Song, K., Giesy, J. P., & Wang, Q. (2020). A novel Mg(OH)2 binding layer-based DGT technique for measuring phosphorus in water and sediment. *Environmental Science: Processes* and Impacts, 22(2), 340–349. https://doi.org/10.1039/c9em00508k
- Xie, J., Wang, Z., Lu, S., Wu, D., Zhang, Z., & Kong, H. (2014). Removal and

recovery of phosphate from water by lanthanum hydroxide materials. *Chemical Engineering Journal*, 254, 163–170.

https://doi.org/10.1016/j.cej.2014.05. 113

- Yabuki, L. N. M., Colaço, C. D., Menegário, A. A., Domingos, R. N., Kiang, C. H., & Pascoaloto, D. (2014). Evaluation of diffusive gradients in thin films technique (DGT) for measuring Al, Cd, Co, Cu, Mn, Ni, and Zn in Amazonian rivers. Environmental Monitoring and 961-969. Assessment. 186(2), https://doi.org/10.1007/s10661-013-3430-x
- Zhang, H., & Davison, W. (1999). Diffusional characteristics of hydrogels used in DGT and DET techniques. *Analytica Chimica Acta*, 398, 329–340.
- Zhang, Y., Song, J., Zhou, H., Zhang,
 Y., & Wang, G. (2018). Novel Fe3O4
 nanoparticles-based DGT device for
 dissolved reactive phosphate
 measurement. New Journal of
 Chemistry, 42(4), 2874–2881.
 https://doi.org/10.1039/c7nj04464j
- Zhang, Z., Yu, H., Zhu, R., Zhang, X., & Yan, L. (2020). Phosphate adsorption performance and mechanisms by nanoporous biochar–iron oxides from

aqueous solutions. *Environmental Science and Pollution Research*, *27*(22), 28132–28145. https://doi.org/10.1007/s11356-020-09166-5

- Zheng, S., Sheng, F., Gu, C., Li, Y., Fang,
 Z., & Luo, J. (2023). DGT method for
 the in situ measurement of triazines
 and the desorption kinetics of atrazine
 in soil. *Environmental Science and Pollution Research*, 30(17), 51061–
 51074.
 https://doi.org/10.1007/s11356-02325985-8
- Q., Maeno, Zhu, S., Sasaki, M., Miyamoto, T., & Fukushima, M. (2015). Monopersulfate oxidation of 2,4,6-tribromophenol using an iron(III)-tetrakis(psulfonatephenyl)porphyrin catalyst supported on an ionic liquid functionalized Fe₃O₄ coated with silica. *B*: Applied Catalysis Environmental. 163. 459-466. https://doi.org/10.1016/j.apcatb.2014. 08.035
 - Zucca, P., Fernandez-Lafuente, R., & Sanjust, E. (2016). Agarose and its derivatives as supports for enzyme immobilization. In *Molecules* (Vol. 21, Issue 11). MDPI AG. https://doi.org/10.3390/molecules211 11577