

WORLD CHEMICAL ENGINEERING JOURNAL

Journal homepage: http://jurnal.untirta.ac.id/index.php/WCEJ

TiO₂/Chitosan bioplastic as Antibacterial of *Stephylococcus aureus* for Food Preservation

Indar Kustiningsih^{1*}, Dhena Ria Barleany¹, Devi Abriyani¹, Asep Ridwan, Muhammad Syairazy, Mochamad Adha Firdaus

¹Chemical Engineering Department Sultan Ageng Tirtayasa University, Cilegon, Indonesia ²Chemical Engineering Department King Fahd University of Petroleum and Minerals, Dhanran, Saudi Arabia

*Corresponding Author Email: indar.kustiningsih@untirta.ac.id

ARTICLE HISTORY	ABSTRACT
Received April 7 th , 2021 Received in revised form May 19 th , 2021 Accepted June 22 th , 2021 Available online September 1 st , 2021	Nowadays, bioplastic development become hot trends to assess environmental issues. Many materials have been purposed to be the best resources for bioplastic manufacturing. Chitosan is one of the most abundant resources in which could derivates from biomaterial waste called chitin. TiO ₂ nanoparticles incorporation within biomaterial presumably not only enhance its mechanical properties but also improve biocompatibility of medical characteristic such as bacterial annihilation. From this study, it was shown that small amount of TiO ₂ nanoparticles within chitosan bioplastic prove improvement of both characteristic. Nevertheless, it was also slightly increasing

material durability to degrade.

Keywords: TiO2, bioplastic, Antibacterial, Stephylococcos aureus

1. INTRODUCTION

Recent development of biodegradable plastic manufacturing for food preservation to also solve environmental issues aimed from second derivatives of natural ingredients like chitosan (Diaz, V. et al., 2010; Fathanah, dkk., 2018; Ginting, dkk., 2018; Naito et al., 2016; Nishiyama et al., 1996; Ogawa, et el., 2019; Rohmawati, dkk., 2018, Kustiningsih, I., et. al., 2019). Chitosan is one of the biggest natural biopolymer sources after cellulose (Mallakpour & Madani, 2015) that have excellent biodegradable features, nontoxic, and biocompatible (Haldorai & Shim, 2014; Mazin C., 2015; Nikkhoo et al., 2018). Chitosan also have antimicrobial properties and easy to produce with low price (Kashif & Park, 2019; Logpriya et al., 2018; Panariello, Coltelli, Buchignani, & Lazzeri, 2019). Compare to conventional plastic, chitosan has a back draw predominantly in its mechanical properties such as tensile strength and elongation (Mallakpour & Madani, 2015)..

Manufacturing nanocomposite chitosan with nanofiller such as silica, clay, and TiO_2 particles predominantly tended to improve not only mechanical

properties but also enhance biomedical features within nanocomposite material specifically for food preservation (de Azeredo, 2009; Kanmani & Rhim, 2014a; Rhim, 2011; Rhim, Park, & Ha, 2013). Nowadays, nanofiller classified into several specific type based on its shape, including: nanoparticles (Kanmani & Rhim, 2014, Kustiningsih, I., et al., 2019.), nanofibril (Rafieian, Shahedi, Keramat, & Simonsen, 2014) and nanotubes ((Diaz-Visurraga et al., 2010). Nanoparticle TiO2 incorporation within chitosan material proven to alter its antimicrobial apart from other nanofiller by growthcontrol and invasion of bacteria that leads to elimination of pathogenic microorganism (de Azeredo, 2009).

TiO₂ has been widely developed for its medical features such as virus, fungi, algae and cancer cell elimination (Chawengkijwanich & Hayata, 2008; Tsuang et al., 2008). Furthermore, TiO₂ proven could annihilates several bacteria (*Pseudomonas aeruainosa* and Enterococcus faecalis bacteria (Jeffery, Peppler, Lima, & McDonald, 2010); Lactobacillus helveticus (Liu & Yang, 2003)). Furthermore, there are few reports on the observation of pure nanoparticles TiO₂ for specified Staphylococcus aureus elimination within chitosan bioplastic. The experiment of manufacturing TiO_2 /chitosan bioplastic and its application specifically for food preservation will leads to the opportunity of advancement bio composite material.

2. METHODS

One sample of 1-gram chitosan mixed with 1% (v/v) acetic acid glacial, stirred for 3 hours and homogenized for 30 minutes at room temperature. Followed by TiO₂ incorporation for each variation (TiO₂: 0 g, 0.1 g, 0.2 g, 0.5 g and 1 g), then stirred for 4 hours and homogenized for 1 hour. A solution molded into glass plate and dried at 80°C afterwards.

The characterization of bioplastic was done using Hitachi SU-3500 Scanning Electron Microscope to assess surface morphology, shape smoothness and TiO₂ particle dispersion. Prestige Shimadzu Fourier Transform Infra-Red 8201 was used for chemical functional groups analysis and Universal Testing Machine to analyze mechanical properties such as elongation and tensile strength.

Sterilized nutrient agar medium has been mixed with sample bioplastic. Followed by blending with rejuvenated Staphyloccus aureus then dissolved with 103 and 104 solution. Thereafter, plating the mixture upon petri dish for 24 hours incubation for different environment: dark and under UV radiation. Bacteria colony counted afterwards by colony forming units (CFU).

3. RESULT and DISCUSSION

3.1. Spectrophotometer FTIR

Fourier Transform Infrared Spectroscopy have observed functional groups analysis involving: O-H bond, N-H bond, C=O bond, C-O bond, and C-N bond for pure chitosan bioplastic sample whereas R-NH-R bond and Cs-TiO₂ were detected for TiO₂/chitosan bioplastic (**Figure 1**).

The graph shows similarity between pure chitosan bioplastic sample and TiO2 incorporated chitosan bioplastic. Characteristics transformation of transmittance peak from FTIR analysis occurred as a result from pure chitosan bioplastic mixed with TiO2 nanoparticles. The mixture has not only physical interaction but also chemical bond (Bourtoom & Chinnan, 2008). Compare to pure chitosan bioplastic, there were shifting of hydroxyl bonds, amino bonds, and amide bonds of TiO2 incorporated chitosan bioplastic. The changes caused by interaction between chitosan molecules and TiO2 nanoparticles (Haldorai & Shim, 2014) which is shown by chitosan – TiO2 bonds on 450 - 950 wavelength range (Amir, Julkapli, & Hamid, 2016).

The spectrum shows that pure chitosan bioplastic has an O-H bonds and N-H bonds at the peak of 3419 cm-1 whereas functional groups of C-H exhibited at 2924 cm-1, C=O at 1616 cm-1, C-O at 1257 cm-1 and C-N at 1039 cm-1 respectively. On the other hand, peak spectrum of TiO2 incorporated chitosan bioplastic is a combination of each pure peak compound's chitosan and TiO2. It could be said with certainty that there was a cross linkage between chitosan and TiO₂ functional groups as for example the overlapped O-H bonds with N-H bonds at 3439 cm-1, R-NH-R at 1600 cm-1, O-H deformation at 1442 cm-1, Chitosan/TiO2 deformation at 947 cm-1.

3.2. Surface layer analysis

The results of Scanning Electron Microscope (SEM) describes the difference upon surface layer of pure chitosan bioplastic and TiO_2 incorporated chitosan bioplastic for each composition (Figure 2). The morphology of pure chitosan bioplastic displays such a homogenous, sleek and clean without any speckles. However, TiO_2 nanoparticles enhancement aimed chitosan bioplastic have a rough structure surface with many grains layered upon.



Figure 1. FTIR spectrum of Cs dan TiO₂/chitosan bioplastic



Fig. 2. . Scanning Electron Microscope (SEM) image of chitosan bioplastic and TiO2 incorporate chitosan bioplastic (a) Cs (b) Cs-OH (c) Cs-O.1 Ti (d) Cs-O.2 Ti (e) Cs-O.3 Ti.

These speckles reveal TiO_2 nanoparticles dispersion within chitosan bioplastic matrix (Figure 2b to Figure 2e). TiO_2 nanoparticles started to exhibit regular pattern when the composition of TiO_2 at 1 : 0.1 within chitosan bioplastic (Figure 2c). At ratio of 1 : 0.2 and 1 : 0.3 (Cs : TiO_2), the distribution of TiO_2 nanoparticles within chitosan bioplastic manifests such a fine cluster compare to the chitosan bioplastic surface (Figure 2d and Figure 2e). A proper distribution of TiO_2 nanoparticles within matrix polymer is one of key indicator of a good TiO_2 incorporated chitosan bioplastic (Diaz-Visurraga et al., 2010).

3.3. Mechanical Properties analysis (Tensile Strength and Elongation)

Tensile strength and elongation tests are involved on mechanical property analysis of chitosan bioplastic and TiO_2 incorporated chitosan bioplastic (Table 2, Figure 3). TiO_2 enhancement on chitosan bioplastic reduce the both of mechanical features. The amount of TiO_2 nanoparticles composition within chitosan bioplastic change the matrix from homogenous into heterogeneous as appears upon surface layer. The phenomena occurred caused by the irregularity of TiO_2 nanoparticles distribution within chitosan bioplastic matrix as shown

in Figure 2b to Figure 2e. These finding also align from others that discovered the effect of TiO_2 nanoparticles on mechanical properties of chitosan (Mazin C., 2015).

Enhancement of TiO₂ nanoparticles within chitosan bioplastic gave a positive results of mechanical properties improvement. For a sample with 0.1 g TiO₂ composition, tensile strength exhibited is 16.93 MPa (decreasing 10.55 MPa) with observed elongation is 8.67 % (increasing 6.02%) from control sample which made from pure chitosan bioplastic. Furthermore, for 0.2 g TiO₂ composition, observed tensile strength is 29.46 (increasing 1.98 MPa) with elongation exhibited was 8.67 % (same with 0.1 g TiO₂).

Afterwards, increasing TiO_2 nanoparticles into chitosan bioplastic mixture seems to be declining its mechanical properties. It was shown that for TiO2 composition at 0.3 g, 1 g, 2 g and 3 g, all of them have smaller mechanical properties rather than control sample of pure chitosan. These phenomena happened due to influence of TiO_2 nanoparticles that made a change in matrix structure of chitosan. Pure chitosan bioplastic has a homogeneous matrix structure compare to TiO2 incorporated chitosan bioplastic (Figure 2a to Figure 2e).



Table 2. Influence of TiO₂ nanoparticles to mechanical properties chitosan bioplastic

Fig. 3 Mechanical properties analysis (a) Tensile strength; (b) Elongation

3.4. Biodegradation and Antibacterial activity analysis

Biodegradation analysis results after 2 months burial is shown in Table 3. TiO_2 enhancement did not inhibit degradation process of chitosan bioplastic. Mass reducing of bioplastic sample still exhibited due to natural ingredient of bioplastic material (Figure 4). Mass reducing showed up because natural ingredient prone to easier to be digested by several microorganism. Thus, the more TiO_2 nanoparticles enhanced within chitosan bioplastic apt to prevent the process that leads to the longer time required for degradation. This antimicrobial activity from TiO_2 nanoparticles is become one of the features that could effectively prolong chitosan bioplastic endurance from aging.

Table 3. Influence of T_1U_2 nanoparticles to mass losses of chitosan bioplastic and T_1U_2 /chito

Sample	Initial weight (g)	Final weight (g)	Mass losses (%)
Cs	0.05	0.02	60.00
Cs-0,1 Ti	0.10	0.05	50.00
Cs-0,2 Ti	0.10	0.05	50.00
Cs-0,3 Ti	0.14	0.09	35071
Cs-0,4 Ti	0.15	0.10	33.33
Cs-0,5 Ti	0.17	0.12	29.41



Figure 4 Biodegradation analysis of chitosan bioplastic and TiO₂/chitosan bioplastic

Photocatalytic TiO₂ by UV irradiation has been done to verify antibacterial bioplastic characteristic in order to annihilate gram-negative bacteria and gram-positive bacteria including: endospore, fungi, algae, protozoa and also virus (Paspaltsis et al., 2006). Staphylococcus aureus has been used as a representation of grampositive bacteria. The results of antibacterial activity presented in Table 4.

As by nature, chitosan bioplastic has an antibacterial activity features but not to eliminate the numbers of Stephylococcus aureus. The antibacterial properties come from amino functional groups that has positive charge which could interact with membrane cell of microorganism which has negative charge. This interaction would cause a destruction of protein and other intracellular constituent from microbes. Chitosan has been tested effectively eradicate gram-negative bacteria rather than gram-positive bacteria (Diaz-Visurraga et al., 2010; Zheng et al., 2000)

It has been proven by these experiment and the results that shown in Table 4, TiO2 nanoparticles enhancement could significantly reduce the amount of

s.auerus bacteria that has been incubated for 24 hours. Interestingly, elimination of bacteria s.aureus could be done either by irradiation or without irradiation by UV lights. These kinds of characteristics also observed in other experiments (Diaz-Visurraga et al., 2010; Tsuang et al., 2008). Photocatalytic activity of TiO2 nanoparticles can be initiated by UV irradiation that will release positive ion charge within chitosan bioplastic. These positive ion charge then will interact with lipid membrane of s.aureus bacteria in which have negative ion.

Under UV irradiation, TiO₂ nanoparticles will generate electron (e-) on conduction band and exhibit the hole (h+) on valence band. The hole will interact with water molecule to produce reactive radical hydroxyl (·OH) and ·O2 which lead to degradation of any organic substance like membrane barrier of cell bacteria (Gumiero et al., 2013). Without UV irradiation, the mechanism of s.aureus bacteria elimination within TiO₂ nanoparticles incorporated mixture remains unknown. It was presumably done by similar process in which Ag elements become an antibacterial agent. Both of Ag and TiO_2 have a positive charge that can interact with negative charge which comes from membrane barrier of cell bacteria (Amrulia, 2012). After 24 hours incubation, the number of control variable bacteria tended to be decreased (Figure 6) for both methods (photocatalytic

and non-photocatalytic). On pure chitosan bioplastic, bacteria only occurred in several colony. In addition, for every sample tested using TiO_2 /chitosan bioplastic there is none of Staphylococcus aureus bacteria prone to be lived.

Sample -	UV irradiation		Without radiation	
	CFU (mL ⁻¹)	Survival Ratio (%)	CFU (mL ⁻¹)	Survival Ratio (%)
Control	2.9 x 10 ⁵	100.0	2.9 x 10 ⁵	100.0
Cs	$2.0 \ge 10^4$	6.9	$2.0 \ge 10^4$	3.5
TiO ₂	0.0	0.0	0.0	0.0
Cs-0,1 Ti	0.0	0.0	0.0	0.0
Cs-0,2 Ti	0.0	0.0	0.0	0.0
Cs-0,3 Ti	0.0	0.0	0.0	0.0
Cs-0,4 Ti	0.0	0.0	0.0	0.0
Cs-0,5 Ti	0.0	0.0	0.0	0.0

Table 4 Antibacterial activity analysis





Figure 5. Antibacteria analysis (a)non photocataltic (b) photocatalytic under UV

4. CONCLUSIONS

 TiO_2 incorporation within chitosan bioplastic is viable to be manufactured. For specified application of food preservation, TiO_2 nanoparticles enhancement exhibits improvement of not only mechanical properties features such as tensile strength and elongation but also able to annihilates the number of Staphylococcus aureus bacteria to zero either by using photocatalytic of nonphotocatalytic method. Nano filling TiO_2 within chitosan bioplastic also made material slightly more durable to degradation compare to conventional plastic.

5. ACKNOWLEDGMENTS

The authors would like to thank to the Islamic Development Bank for financial assistance under IDB-UNTIRTA Research Grant project No. 593/UN43.9/PL/2019.

6. REFERENCES

- Amir, M. N. I., Julkapli, N. M., & Hamid, S. B. A. (2016). Incorporation of chitosan and glass substrate for improvement in adsorption, separation, and stability of TiO2 photodegradation. International Journal of Environmental Science and Technology, 13(3), 865-874. doi:10.1007/s13762-015-0914-y
- Amrulia, W. (2012). Uji Aktivitas Antibakteri Kitosan-TiO₂ Pada Tekstil Terhadap Eschericia coli. [Antibacteria activity test of chitosan -TiO₂ on textile material for Eschericia coli].
- Bourtoom, T., & Chinnan, M. S. (2008). Preparation and properties of rice starch-chitosan blend biodegradable film. Lwt-Food Science and Technology, 41(9), 1633-1641. doi:10.1016/j.lwt.2007.10.014
- Cai, R. X., Kubota, Y., Shuin, T., Sakai, H., Hashimoto, K., & Fujishima, A. (1992). Induction of Cytotoxicity by Photoexcited Tio2 Particles. Cancer Research, 52(8), 2346-2348. Retrieved from <Go to ISI>://WOS:A1992HN84200036
- de Azeredo, H. M. C. (2009). Nanocomposites for food packaging applications. Food Research International, 42(9), 1240-1253. doi:10.1016/j.foodres.2009.03.019
- Diaz-Visurraga, J., Melendrez, M. F., Garcia, A., Paulraj, M., & Cardenas, G. (2010). Semitransparent Chitosan-TiO2 Nanotubes Composite Film for Food Package Applications. Journal of Applied Polymer Science, 116(6), 3503-3515. doi:10.1002/app.31881
- Dompeipen, E. J. (2017). Isolasi dan identifikasi kitim dan kitosan dari kulit udang windu (Penaeus monodon) dengan spektroskopi inframerah, Majalah BIAM, (2017), 12.
- Fathanah, U., Lubis, M. R., Nasution, F., & Masyawi, M. S. (2018). Characterization of bioplastic based from cassava crisp home industrial waste incorporated with chitosan and liquid smoke. 3rd International Conference on Chemical Engineering Sciences and Applications 2017 (3rd Icchesa 2017), 334. doi:Unsp 012073
- 10.1088/1757-899x/334/1/012073
- Ginting, M. H. S., Lubis, M., Sidabutar, T., & Sirait, T. P. (2018). The effect of increasing chitosan on the characteristics of bioplastic from starch talas (Colocasia esculenta) using plasticizer sorbitol. Friendly City 4 from Research to Implementation for Better Sustainability, 126. doi:Unsp 012147

10.1088/1755-1315/126/1/012147

- Gumiero, M., Peressini, D., Pizzariello, A., Sensidoni, A., Iacumin, L., Comi, G., & Toniolo, R. (2013). Effect of TiO₂ photocatalytic activity in a HDPE-based food packaging on the structural and microbiological stability of a short-ripened cheese. Food Chemistry, 138(2-3), 1633-1640. doi:10.1016/j.foodchem.2012.10.139
- Haldorai, Y., & Shim, J. J. (2014). Novel Chitosan-TiO2 Nanohybrid: Preparation, Characterization, Antibacterial, and Photocatalytic Properties. Polymer Composites, 35(2), 327-333. doi:10.1002/pc.22665
- Jeffery, B., Peppler, M., Lima, R. S., & McDonald, A. (2010). Bactericidal Effects of HVOF-Sprayed Nanostructured TiO2 on Pseudomonas aeruginosa. Journal of Thermal Spray Technology, 19(1-2), 344-349. doi:10.1007/s11666-009-9369-3
- Kanmani, P., & Rhim, J. W. (2014a). Physicochemical properties of gelatin/silver nanoparticle antimicrobial composite films. Food Chemistry, 148, 162-169. doi:10.1016/j.foodchem.2013.10.047
- Kanmani, P., & Rhim, J. W. (2014b). Properties and characterization of bionanocomposite films prepared with various biopolymers and ZnO nanoparticles. Carbohydrate Polymers, 106, 190-199. doi:10.1016/j.carbpol.2014.02.007

- Kashif, S. A., & Park, J. K. (2019). Enzymatically Hydrolyzed Water-Soluble Chitosan as a Potent Anti-Microbial Agent. Macromolecular Research, 27(6), 551-557. doi:10.1007/s13233-019-7095-3
- Kustiningsih I, Ridwan A, Abriyani D, Syairazy M, Kurniawan T, Barleany D. R. Development of Chitosan-TiO2 Nanocomposite for Packaging Film and its Ability to Inactive Staphylococcus Aureus. Orient J Chem 2019;35(3). Available from: https://bit.ly/2MUXwF1
- Liu, H. L., & Yang, T. C. K. (2003). Photocatalytic inactivation of Escherichia coli and Lactobacillus helveticus by ZnO and TiO2 activated with ultraviolet light. Process Biochemistry, 39(4), 475-481. doi:10.1016/S0032-9592(03)00084-0
- Logpriya, S., Bhuvaneshwari, V., Vaidehi, D., SenthilKumar, R. P., Malar, R. S. N., Sheetal, B. P., . . . Kalaiselvi, M. (2018). Preparation and characterization of ascorbic acid-mediated chitosan-copper oxide nanocomposite for anti-microbial, sporicidal and biofilminhibitory activity. Journal of Nanostructure in Chemistry, 8(3), 301-309. doi:10.1007/s40097-018-0273-6
- Mallakpour, S., & Madani, M. (2015). Effect of Functionalized TiO2 on Mechanical, Thermal and Swelling Properties of Chitosan-Based Nanocomposite Films. Polymer-Plastics Technology and Engineering, 54(10), 1035-1042. doi:10.1080/03602559.2014.974194
- Mazin C., T., A., Anandapadmanabhan, Ashfaq, Mujeeb, A., dan Lobo, A. G. (2015). Study on the Effect of Nano TiO2 on Mechanical Properties of Chitosan. IOSR Journal of Mechanical and Civil Engineering, 12(3), 7. doi:10.9790/1684-12314854
- Naito, P. K., Ogawa, Y., Sawada, D., Nishiyama, Y., Iwata, T., & Wada, M. (2016). X-ray Crystal Structure of Anhydrous Chitosan at Atomic Resolution. Biopolymers, 105(7), 361-368. doi:10.1002/bip.22818
- Nikkhoo, M., Amini, M., Farnia, S. M. F., Mandavinia, G. R., Gautam, S., & Chae, K. H. (2018). Preparation and Characterization of Magnetic Chitosan/Cu-Mg-Al Layered Double Hydroxide Nanocomposite for the One-Pot Three-Component (A(3)) Coupling of Aldehydes, Amines and Alkynes. Journal of Inorganic and Organometallic Polymers and Materials, 28(5), 2028-2035. doi:10.1007/s10904-018-0861-4
- Nishiyama, M., Hosokawa, J., Yoshihara, K., Kubo, T., Kabeya, H., Endo, T., & Kitagawa, R. (1996). Biodegradable plastics derived from cellulose fiber and chitosan. Hydrophilic Polymers, 248, 113-123. Retrieved from <Go to ISI>://WOS:A1996BE65L00007
- Ogawa, Y., Naito, P. K., & Nishiyama, Y. (2019). Hydrogen-bonding network in anhydrous chitosan from neutron crystallography and periodic density functional theory calculations. Carbohydrate Polymers, 207, 211-217. doi:10.1016/j.carbpol.2018.11.042
- Panariello, L., Coltelli, M. B., Buchignani, M., & Lazzeri, A. (2019). Chitosan and nano-structured chitin for biobased anti-microbial treatments onto cellulose based materials. European Polymer Journal, 113, 328-339. doi:10.1016/j.eurpolymj.2019.02.004
- Paspaltsis, I., Kotta, K., Lagoudaki, R., Grigoriadis, N., Poulios, I., & Sklaviadis, T. (2006). Titanium dioxide photocatalytic inactivation of prions. Journal of General Virology, 87, 3125-3130. doi:10.1099/vir.0.81746-0
- Rafieian, F., Shahedi, M., Keramat, J., & Simonsen, J. (2014). Mechanical, thermal and barrier properties of nano-biocomposite based on gluten and carboxylated cellulose nanocrystals. Industrial Crops and Products, 53, 282-288. doi:10.1016/j.indcrop.2013.12.016
- Rhim, J. W. (2011). Effect of clay contents on mechanical and water vapor barrier properties of agar-based nanocomposite films. Carbohydrate Polymers, 86(2), 691-699. doi:10.1016/j.carbpol.2011.05.010
- Rhim, J. W., Park, H. M., & Ha, C. S. (2013). Bio-nanocomposites for food packaging applications. Progress in Polymer Science, 38(10-11), 1629-1652. doi:10.1016/j.progpolymsci.2013.05.008
- Rohmawati, B., Sya'idah, F. A. N., Rhismayanti, Alighiri, D., & Eden, W. T. (2018). Synthesis of Bioplastic-based Renewable Cellulose Acetate from Teak Wood (Tectona grandis) Biowaste Using Glycerol-Chitosan Plasticizer. Oriental Journal of Chemistry, 34(4), 1810-1816. doi:10.13005/ojc/3404014
- Tsuang, Y. H., Sun, J. S., Huang, Y. C., Lu, C. H., Chang, W. H. S., & Wang, C. C. (2008). Studies of photokilling of bacteria using titanium dioxide nanoparticles. Artificial Organs, 32(2), 167-174. doi:10.1111/j.1525-1594.2007.00530.x
- Zheng, H., Maness, P. C., Blake, D. M., Wolfrum, E. J., Smolinski, S. L., & Jacoby, W. A. (2000). Bactericidal mode of titanium dioxide photocatalysis. Journal of Photochemistry and Photobiology a-

Chemistry, 130(2-3), 163-170. doi:Doi 10.1016/S1010-6030(99)00205-1