EFFECTS OF WASHING TREATMENT ON THE CHARACTERISTICS OF

ROADSIDE FRESH FRUITS

Julfi Restu Amelia, N. Ambarsari, Giyatmi Giyatmi* Department of Food Technology, Sahid University, Indonesia

*E-mail: giyatmi@hotmail.com

ABSTRACT

Roadside fruits are very possible to be contaminated by heavy metals and microbial contamination, especially fruits that are usually consumed unpeeled, such as grapes, water guava, apples, and starfruit. Heavy metal contaminants that are hazardous to health include lead (Pd), cadmium (Cd), and arsenic (As), while harmful microbial contaminants include *Escherichia coli* and *Salmonella thypii*. This study aims to determine the effect of washing treatment on the characteristics (heavy metal and microbial contamination) of some fresh fruit. Washing process using running water and food grade soap. The design of the study used is Randomized Block Design, if it had a significant effect, it was further tested using the Duncan Multiple Range Test (DMRT). The results of study showed that washing with food grade soap is preferable, as it shows a significant reduction in the levels of heavy metal and microbiological contents up to 0.000 mg / kg for Pb and Cd; 0.041 mg / kg for As; 0 kol / g for E. coli and 0 kol / 25g for *Salmonella thypii*.

Keywords: food grade soap, Fruits, heavy metal, microbial contamination

INTRODUCTION

Fruits are an essential part of the human diet. They are sources of many essential nutrients, vitamins, minerals, have high water and fiber content. Fruits that can be directly consumed without needing to be peeled such as grapes, apples, water guava, star fruit, and many others. Farmers or sellers often sell fruits at roadside stands. Road side fruits are potentially exposed to exhaust gases or vehicle fumes that contain heavy metal contamination. Lead, cadmium, and arsenic are among the most risky heavy metals when consumed through contaminated fruits. Of all the heavy metals, cadmium and lead have more significant side effects on human health since they are easily accessible through food chain transmission (Rahimi, 2013). Consumption of roadside

fruits also potentially increases the risk of foodborne illness caused by microbial contaminants. The common microbial contaminants are Esceherichia coli and Salmonella thypii. Another way to reduce heavy metal contamination and microbiology in roadside fruits is washing treatment. Most people wash fruits only by using water. However, this has not guaranteed the contamination of heavy metals and microbiology lost from fruits. Nowadays there are several food grade soap products that contain anionic and antibacterial agent surfactants that are safe to use to wash fresh foods.

This study aimed to measure the effectiveness of washing treatment for reduce the presence of heavy metals and microbiology contamination in road side fruits. The study was carried out using fruits that were purchased directly without washing, compared to washed fruits with running water, and washed fruits using food grade soap. The samples used for research are fruits that are usually consumed directly without peeling, such as grapes, apples, water guava, and star fruit. The results of this study are expected to provide information to the public about the dangers of heavy metals and microbiological contamination of fruits, and can educate the public to do proper washing of fruits, especially fruits that are usually consumed directly without peeling.

MATERIALS AND METHODS Materials

This study uses four types of samples, there are red grapes, Washington apple, Semarang water guava, and Dewi starfruit. Red grapes and Washington apples represent imported fruits while Semarang guava fruit and Dewi star fruit represent local fruits. The fruits used in the study were purchased in the same place, which is in the stalls along the roadside of Pulo Jahe market, East Jakarta. The washing solution used is food grade soap. The active ingredients used in food grade soaps are anionic surfactants and antibacterial food grade agents that can clean pesticides and kill germs up to 99%. The concentration of washing solution used for fruit washing was also the same for each type of fruit, the formulation is 1/2 tbsp (7.5 ml) dissolved in 1 liter of water.

Materials used for testing include, aquadest, acidic solution, Pb standard solution; CD; and As, Modified Letheen Broth media; Nutrient Agar and Sabuoraud Dextrose Agar; Cromocult Agar; and Xylose Lysine Deoxycholate. The tools used for testing included a basin, hollow and plastic basin, beaker glass, stirring rod, hot plate, measuring flask, glass funnel, filter paper, Atomic Absorption Spectrophotometry (AAS), Gas Chromatography (GC), magnetic stirrer, bottles sterile, test tube, finnpipette, pipette tips, autoclave, Bio Safety Cabinet (BSC), and incubator.

Procedure

The steps of determining the effect of washing on heavy metal and microbiological contamination on fresh fruit can be seen in Figure 1 and Figure 2. In the first step, the fruits analyzed were selected through screening results including the selection of all types of fruits that can be consumed without peeling the skin that are sold in fruit stalls along the Pulo Jahe market road using descriptive methods. The microbiology test carried out was the total calculation of microbes including bacteria, fungi, and yeast using the Total Plate Count (TPC) method. While the chemical test using the Gas Chromatography (GC) method.

Chemical Test Heavy Metal Contents

Fruit samples prepared before reading with AAS instrument. The fruits were initially deconstructed, which was weighed as much as 100 grams in a porcelain dish, burned, and ashed in a furnace at 900°C for 1 hour. Then the ash filtrate was acidified with the addition of 1 ml of 1: 1 HCl and being heated. After dissolving, put the solution into a 100 ml measuring flask and add aquadest to the boundary mark. The solution is filtered and ready to read the content of heavy metals with the AAS instrument at the wavelength specified for each heavy metal.

Moisture Content (AOAC, 2006)

Weighed each dish that will be used. weighed ± 5 grams of sample into the dish, and put the dish in the oven at 105°C for ± 4 hours or until the weight is constant, then cooled in the desiccator, and weighed at the final dish with its sample. The calculation of wet and dry base water content was used. The formula for calculating water content is as follows:

> Moisture content (wet base) (%) = $\frac{initial weight - final weight}{initial weight} x 100\%$ Moisture content (dry base) (%) = $\frac{initial weight - final weight}{final weight} x 100\%$

Ash Content (AOAC, 2006)

Prepare the porcelain dish to do the obfuscation, then dry it in an oven for 15 minutes and then cool it in a desiccator and weighed (A). The sample is weighed ± 3 grams in a porcelain dish (B), then it is burned in chamber until it doesn't smoke again. Then put it in electric furnace at a temperature of $400 - 600^{\circ}$ C for 4 - 6 hours until white ash was formed or had a fixed weight. The ash and cup are cooled in a desiccator then weighed (C).

Ash Content (% bb) = $\frac{(C - A)}{B} \times 100 \%$

Protein Content (AOAC, 2006)

Samples of \pm 100 mg were weighed (A) and put into a 30 ml kjeldahl flask. Then added 1.9 \pm 0.1 gr K₂SO₄, 40 \pm 10 mg HgO, and 3.8 \pm 0.1 ml H₂SO₄. Boiling stone is added to the flask and the sample is boiled for 1-1.5 hours until the liquid becomes clear. The contents of the flask and the rinsed water are transferred into a distillation instrument. The 125 ml erlenmeyer flask was filled with 5 ml of H_3BO_4 solution and added with 4 drops of indicators, then placed under a condenser with the tip of the condenser submerged in either H_3BO_4 solution. 8-10 ml NaOH-Na₂S₂O₃ solution was added to the distillation apparatus until it obtained distillate \pm 15 ml in erlenmeyer. The distillate in the erlenmeyer is then titrated with 0.02 N HCl solution until there is a green to blue color change. Calculated the amount of nitrogen after previously obtained the amount of volume (ml) blank.

Amount of N (%) =

 $\frac{(\mathsf{mI}\;\mathsf{HCI}-\mathsf{mI}\;\mathsf{blank}) \times \mathsf{N}\;\mathsf{HCI}\times \mathsf{14.007}}{\mathsf{A}}\;\;x\;\;100\;\;\%$

Fat Content (AOAC, 2006)

Fat flask is provided according to the size of the soxhlet extraction tool used. The flask was dried in an oven at a temperature of 105 - 110 oC for 15 minutes, then cooled in a desiccator then weighed (A). Weighed as much as ± 5 g of sample (B) in filter paper, then covered with fat-free cotton. Filter paper and its contents are put into the Soxhlet extraction and installed on the condenser. The hexane solvent is poured into the Soxhlet flask to taste. Then it is refluxed for 5 hours until the solvent that comes back becomes clear. The solvent remaining in the fat flask is distilled and then the flask is heated in an oven at 105oC. After being dried to a fixed weight and cooled in a desiccator, the flask and the fat are weighed (C) and the following fat content is calculated.

Fat content (%) =
$$\frac{C-A}{B} \times 100 \%$$

Carbohydrate Content by difference (AOAC, 2006)

Carbohydrate content is calculated based on by difference method with the following calculation:

Carbohydrate content (% ww) = 100% -(moisture content + ash content + protein content + fat content)

Microbiological Test Escherichia coli Identification

Microbiological testing on fruits was carried out by quantitative identification of E. coli-Coliform. Where fruit samples are weighed 1 gram and dissolved in media (Modified Letheen Broth) 9ml MLB in aseptic conditions. Then 1 ml was taken to be planted on selective media for E. Coli-Coliform that is Cromocult Agar (CA). After it being solid, incubated at 36°C for 2x24 hours. Positive *E. coli* samples will grow purple colonies, and Coliform positive samples will grow red colonies.

Salmonella thypii Identification

Microbiological testing of fruits for the identification of *S. thypii* was carried out qualitatively. Fruit samples were weighed 25 grams and dissolved in media (Modified Letheen Broth) 225 ml in aseptic conditions. Then incubated at 36° C for 24 hours, to enrich microbes. Then 1 ml was taken to be planted on selective media for *S. thypii* that is Xylose Lysine Deoxycholate (XLD). After it being solid, it was incubated at 36° C for 2x24 hours. Positive samples of *S. thypii* will change the color of the media from brown to red and grow black colonies.

Statistical Analysis

The analysis technique used in this study is variance analysis (ANOVA), using the Statistical Package for Social Science (SPSS) application program. The design of the study used is Randomized Block Design, if it had a significant effect, it was further tested using the Duncan Multiple Range Test (DMRT).

RESULTS AND DISCUSSION Fruits Screening

Roadside fruits may become contaminated with pathogenic microorganisms either during their growing in fields or greenhouses, or during harvesting, postharvest handling, and distribution (Beuchat, 2002).

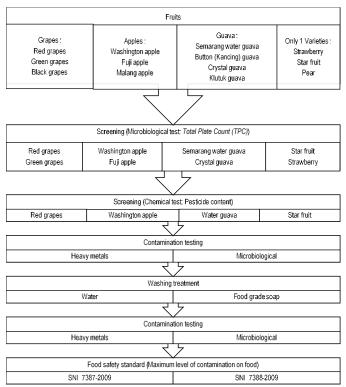


Figure 1. The steps of determining the effect of washing on heavy metal and microbiological contamination on fresh fruit

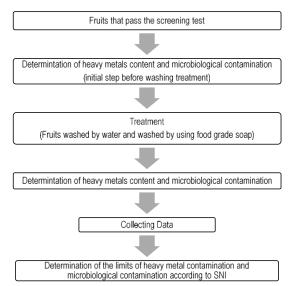


Figure 2. Flow Chart of determining the effect of washing on heavy metal and microbiological contamination on fresh fruit process

Postharvest sources of contamination include faeces, harvesting equipment, human handling, insects, wild and domestic animals, methods of transportation, processing equipment dust, and rinse water (Gil, et al., 2015). The result from fruits screening in microbilogical testing and chemical testing can be seen on Table 1 and Table 2. Table 1 and 2 showed that all roadside fruits showed bacterial, fungi, and yeast contamination, the most contaminated was grapes (red grapes and green grapes). Fruits like grapes, apple, guava, etc are mostly handled with bare hands during harvesting, packaging, and

distribution; hence, it has been established that many fruits in the country are contaminated before they are sold, which, when not properly washed, will result in food-borne illnesses (Boateng, 2016). Each of the fruit varieties with the highest results of chemical analysis is selected one type that will proceed to the main study to determine presence of heavy metal and the microbiological contamination as well as the effect of washing on the decrease in content of heavy metal and microbiological contamination in fruits.

No	Varieties	Sample	Total Plate	Count (TPC) o	colony/gram
140	varieties	Sample	Bacteria	Fungi	Yeast
1	_	Red grapes	9.8 x10 ⁴	$4.2 \text{ x} 10^3$	$1.0 \text{ x} 10^4$
2	Grapes Vitis vinera	Green grapes	$7.4 \text{ x} 10^3$	$<1x10^{1}$	$5.0 \text{ x} 10^3$
3	vitis villeru	Black grapes	$1.5 \text{ x} 10^3$	$<1x10^{1}$	$1.1 \text{ x} 10^2$
4	Angle	Washington apple	$3.0 \text{ x} 10^4$	$4.8 \text{ x} 10^3$	$5.8 \text{ x} 10^3$
5	Apple Malus domestica	Fuji apple	$1.6 \text{ x} 10^4$	$1.8 \text{ x} 10^3$	$1.3 \text{ x} 10^3$
6	Maras domostica	Malang apple	$4.7 \text{ x} 10^3$	$3.0 \text{ x} 10^2$	$2.5 \text{ x} 10^3$
7	Water guava	Semarang water guava	$2.2 \text{ x} 10^5$	$1.3 \text{ x} 10^3$	$2.5 \text{ x} 10^3$
8	Syzygium aqueum	Button guava (Jambu kancing)	1.1 x10 ⁵	$<1x10^{1}$	<1x10 ¹
9	Guava	Crystal guava	1.8 x10 ⁵	2.3 x10 ³	$1.2 \text{ x} 10^3$
10	Psidium guajava	Klutuk guava	$1.4 \text{ x} 10^4$	$2.0 \text{ x} 10^3$	$1.2 \text{ x} 10^3$
11		Strawberry	$2.4 \text{ x} 10^4$	5.3 x10 ³	2.5 x10 ³
12	Only 1 variety	Dewi star fruit	2.9 x10 ⁴	1.1 x10 ⁴	$5.7 \text{ x} 10^3$
13		Pear	$1.4 \text{ x} 10^4$	$2.0 \text{ x} 10^3$	$1.2 \text{ x} 10^3$

Table 1. Result from fruits screening in microbiological testing

Table 2. Result from fruits screening in chemical testing

No	Varieties	Sample	Result (mg/kg)
1	Granas	Red grapes	0.90
2	Grapes	Green grapes	0.43
3	Annla	Washington Apple	0.33
4	Apple	Fuji apple	0.16
5	Guava	Semarang water guava	0,07
6	Guava	Crystal guava	0,03
7	Only 1 variant	Strawberry	0,03
8	Only 1 variant	Dewi star fruit	0,06

Heavy Metals Content

The results of the content of heavy metals analysis for Pb, Cd, and As from each fruit can be seen in the Table 3. Test results of heavy metal content in fruits can be seen from Table 3 have decreased. Test results of variance Pb heavy metal content in fruits showed that washing treatment (treatment significance = 0.204) did not significantly affect Pb content at the level of significance (α) 0.05. PB is very dangerous for the environment. The period of stay of Pb particles in the air is 4-40 days. Pb particles can be spread by the wind to reach distances of 100-1000 km from the source (Ayu, 2000). While Cd heavy metal, the test results of variance Cd heavy metal content showed that washing treatment (treatment significance = 0.371) did not significantly affect the levels of Cd heavy metals at the level of significance (α) 0.05. Cd is a toxic pollutant in water from metal mining, melting, plating, batteries, pesticides, oil paint, pigments, and alloys (Li et al., 2007 and Chen et al., 2008). Large dosages of Cd in humans can cause proteinurea and anemia (Horsfall et al., 2005).

Arsenic (As) is one of the major global environmental pollutants because of its

highly toxic and carcinogenic properties (Bhattacharya, et al., 2010). The epidemiological studies show that the chronic As poisoning can cause cancers, diabetes mellitus (DM), hypertension, melanosis, hyperkeratosis, restrictive lung disease, peripheral vascular, and ischemic heart disease (Mazumder et al. 2000; Morales et al. 2000; Srivastava et al. 2001; Rahman 2002). While the test results of heavy metal content of As in fruits showed that washing treatment (significance of treatment = 0.020) significantly affected the content of As heavy metal at the level of significance (α) 0.05. Further testing in the form of Duncan test was carried out to see the effect of washing treatment on As content in the fruit group. The results of the test showed that the decrease in As content in fruits can be seen in Table 3. As the levels at the beginning before washing had a significant difference to the As content after washing using water only and food grade soap. The level of As in the fruit group after washing using water only and food grade soap has no real difference. This is because As has a difficultity to dissolve in water.

Heavy metals	Washing treatment	Grapes	Apple	Guava	Star fruit
	Initiate	3,973ª	0,216 ^a	0,066ª	1,027ª
Pb	Water treatment	1,390 ^a	0,053ª	$0,000^{a}$	0,403 ^a
PO	Food grade soap treatment	0,000ª	0,000ª	0,000ª	0,023 ^a
	Initiate	1,177ª	0,053ª	$0,000^{a}$	0,037ª
Cd	Water treatment	0,006 ^a	$0,000^{a}$	$0,000^{a}$	0,034ª
Cu	Food grade soap treatment	0,000ª	0,000ª	0,000ª	0,000 ^a
	Initiate	0,141ª	0,141ª	0,243ª	0,074 ^a
1 0	Water treatment	0,089 ^b	0,080 ^b	0,103 ^b	0,064 ^b
As	Food grade soap treatment	0,065 ^b	0,041 ^b	0,085 ^b	0,053 ^b

Table 3. Result of heavy metals analysis (mg/kg) on fruits

Microbiological qualities

The results of microbiological analysis of the identification of E. Coli-Coliform and Salmonella thypii can be seen in Table 4. The decrease in the number of colonies of E. colicoliform and S. thypii due to washing treatment on fruits can be seen in Table 4. From three washing treatments used, it can be seen that washing treatment using food grade soap is better than washing using only with water to decrease the amount of E. colicoliform and S. thypii colonies in the fruit group used. The results of the variance in the identification of E. Coli-Coliform in fruits showed that washing treatment (treatment significance = 0.127) did not significantly influence the decrease in the number of E. Coli-Coliform colonies at the level of significance (α) 0.05. While the results of the identification test for fruit S. Thypii showed

washing treatment (significance = 0.000) significantly affected the decrease in the number of S. thypii at the level of significance (α) 0.05. Further testing in the form of Duncan test was conducted to see the effect of washing treatment on the number of S. thypii in the fruit group. The results of duncan test on the decrease of S. thypii in fruits can be seen in Table 4. It can be seen from the Duncan test results that the number of S. thypii at the beginning before washing and after washing using running water has no real difference, but both have real differences on the number of *S. thypii* after washing using food grade soap. This can be caused by the small number of S. thypii in fruits, so that antibacterial substances in food grade soaps are very effective for eliminating S. thypii.

Table 4. Result of microbiological identification (col/g) on fruits

Microbes	Washing Treatment	Grapes	Apple	Guava	Star fruit
	Initiate	$1x10^{2}$	$3,2x10^{2}$	$5,4x10^4$	$4,4x10^4$
E. coli	Water treatment	3x10 ¹	1,000	1,8x10 ⁴	6,0x10 ³
<i>E. con</i>	Food grade soap treatment	0,000	0,000	8,8x10 ²	3x10 ²
	Initiate	1ª (+)	1ª (+)	1ª (+)	1ª (+)
S thunii	Water treatment	1ª (+)	1ª (+)	1ª (+)	1ª (+)
S. thypii	Food grade soap treatment	0 ^b (-)	0 ^b (-)	0 ^b (-)	0 ^b (-)

Supporting Test

Supporting tests on fruits were carried out to determine the nutritional quality of fruits and to determine the content of active ingredients and antibacterial substances in food grade soaps that have been tested. Proximate testing results and mineral content testing in fruits can be seen in Table 5. The results of the active ingredients content in food grade soap can be seen in Table 6. And the results of antibacterial substances testing on food grade soap can be seen in Table 7.

CONCLUSION

Chemical tests results showed that there was an effect ($<\alpha = 0.05$) of washing treatment on heavy metal contamination in fresh fruits which are usually consumed directly without peeling the skin, and has a significant effect on Arsenic (As) heavy content. For the results metal of microbiological tests showed that there was an effect ($<\alpha = 0.05$) of washing treatment on microbes in fresh fruit which is usually consumed directly without peeling the skin. And significantly affect the pathogenic microbes of Salmonella thypii. Based on the results of this study on fruits washing

treatment using food grade soap which is usually consumed without peeling the skin and sold along the road is more effective in reducing heavy metal and microbiological contamination.

Devenuetor	Result				
Parameter	Grapes	Apple	Guava	Star fruit	
Water content (%)	83.99	83,39	93,77	92,99	
Ash content (%)	0,17	0,01	0,02	0,02	
Fat content (%)	0.09	0.07	0.06	0,07	
Protein content (%)	1,82	1,66	0,89	1,23	
Carbohydrate content (%)	13,15	14,39	5,98	5,49	
Fe (mg/100g)	0,326	0,132	0,073	0,077	
Ca (mg/100g)	10,895	6,774	29,253	3,346	
Mg (mg/100g)	7,141	5,116	5,586	10,583	

Table 5	Result test	of	proximate and	l mineral	content in fruits
Table J	. Result test	01	proximate and	1 mmerai	content in nuits

Table 6. Result test of active agent (Anionic surfactant) in food

Parameter	Result (%)
Lauryl Alkyl Sulphonate Sodium (LAS Na)	10,85
Sodium Lauryl Ethoxy Sulphonate (SLES)	11,17

Table 7. Result test of active agent (A	(Anionic surfactant) in food
---	------------------------------

Parameter	Result (%)
Triclosan	0
TCC	0
Methylisothializone	0
Methylchloroisothializone	0
Zinc Sulphate	1,06

REFERENCES

- Association of Official Analytical Chemists, & Association of Official Agricultural Chemists (US). 2006. Official methods of analysis of the Association of Official Analytical Chemists (Vol. 2). The Association.
- Ayu CC. 2002. Mempelajari Kadar Mineral dan Logam Berat pada Komoditi Sayuran Segar di Beberapa Pasar di Bogor [Skripsi]. Bogor : Fakultas Teknologi Pertanian, Institut Pertanian Bogor.
- Bhattacharya P, Samal AC, Majumdar J, dan Santra SC. 2010. Arsenic contamination in rice, wheat, pulses, and vegetables: a study in an arsenic

affected area of West Bengal, India. Water, Air, & Soil Pollution, 213(1-4), 3-13.

- Beuchat LR. 2002. Ecological factors influencing survival and growth of human pathogens on raw fruits and vegetables. Microb Infect 4: 413–23.
- Boateng CN. 2016. Analysis of Post Harvest Losses in the Mango Marketing Channel in Southern Ghana (Doctoral dissertation, University of Ghana).
- Chen G, Zeng G, Tang L, Du C, Jiang X, Huang G, ... & Shen G. 2008. Cadmium removal from simulated wastewater to biomass byproduct of Lentinus edodes. Bioresource technology, 99(15), 7034-7040.

- Horsfall M, & Spiff AI. 2005. Effect of metal ion concentration on the biosorption of Pb2+ and Cd2+ by Caladium bicolor (wild cocoyam). African Journal of Biotechnology, 4(2), 191-196.
- Gil MI, Selma MV, Suslow T, Jacxsens L, Uyttendaele M, & Allende A. 2015. Pre-and postharvest preventive measures and intervention strategies to control microbial food safety hazards of fresh leafy vegetables. Critical reviews in food science and nutrition, 55(4), 453-468.
- Mazumder DNG, Haque R, Ghosh N, De BK, Santra A, Chakraborti D, & Smith AH. 2000. Arsenic in drinking water and the prevalence of respiratory effects in West Bengal, India. International journal of epidemiology, 29(6), 1047-1052.
- Morales KH, Ryan L, Kuo TL, Wu MM, & Chen CJ. 2000. Risk of internal cancers from arsenic in drinking water. Environmental health perspectives, 108(7), 655-661.
- Rahman M. 2002. Arsenic and contamination of drinking-water in Bangladesh: a public-health perspective. Journal of Health, Population and Nutrition, 193-197.
- Rahimi E. 2013. Lead and cadmium concentrations in goat, cow, sheep, and buffalo milks from different regions of Iran. Food chemistry, 136(2), 389-391.
- Srivastava AK, Hasan SK, & Srivastava RC. 2001. Arsenicism in India: dermal lesions and hair levels. Archives of environmental health, 56(6), 562.
- Li X, Tang Y, Xuan Z, Liu Y, & Luo F. 2007. Study on the preparation of orange peel cellulose adsorbents and biosorption of Cd2+ from aqueous solution. Separation and Purification Technology, 55(1), 69-75.